

ISSN 1669-5402 (Print)

ISSN 1669-5410 (Online)



*Physiological  
Mini-  
Reviews*

*Edited by the Argentine Physiological Society.*

*Vol. 3, N° 2, October-November 2007.*

<http://www.mini.reviews.safisiol.org.ar>

# Physiological Mini-Reviews

[ISSN 1669-5402 (Print); ISSN 1669-5410 (Online)]

Edited by the **Argentine Physiological Society**

Journal address: Sociedad Argentina de Fisiología, Universidad Favaloro, Solís 453 (1078),  
Ciudad de Buenos Aires Argentina.  
Tel.-Fax: (54) (0)11 43781151  
<http://www.mini.reviews.safisiol.org.ar>

---

*Physiological Mini-Reviews* is a scientific journal, publishing brief reviews on "hot" topics in Physiology. The scope is quite broad, going from "Molecular Physiology" to "Integrated Physiological Systems". As indicated by our title it is not our intention to publish exhaustive and complete reviews. We ask to the authors concise and updated descriptions of the "state of the art" in a specific topic. Innovative and thought-provoking ideas are welcome.

---

## **Editorial Board:**

Eduardo Arzt, Buenos Aires, Argentina.  
Oscar Candia, New York, United States.  
Daniel Cardinali, Buenos Aires, Argentina.  
Hugo Carrer, Córdoba, Argentina.  
Marcelino Cerejido, México City, México.  
Horacio Cingolani, La Plata, Argentina.

Adolfo De Bold, Ottawa, Canada.  
Osvaldo Delbono, Salem, United States.  
Cecilia Hidalgo, Santiago, Chile.  
Carlos Libertun, Buenos Aires, Argentina.  
Gerhard Malnic, Sao Paulo, Brasil.  
Raúl Marinelli, Rosario, Argentina.  
Juan Saavedra, Bethesda, United States.  
David Sabatini, New York, United States.

**Editor in Chief:** Mario Parisi.

---

**Annual suscriptions rates** are (see the electronic version for payment instructions):

- a) Printed (Institutions): 120 U\$\$ (Air mail.)
  - b) Printed (Individuals): 100 U\$\$ (Air mail. Including Safis Annual fee.)
  - c) Electronic (Individuals-.PDF): 30 U\$\$ (Including Safis Annual fee.)
  - d) Electronic (Institutions-.PDF): 50 U\$\$
- 

## **Preparation and Submission of manuscripts:**

"Physiological Mini-Reviews" will have a maximum of 2500 words, 30 references and 4 figures. Material will be addressed to scientific people in general but not restricted to specialist of the field. For citations in the text and reference list see Cerejido et al. Vol 1, N° 1. Final format will be given at the Editorial Office. Most contributions will be invited ones, but spontaneous presentations are welcome. Send your manuscript in Word format (.doc) to:  
[mini-reviews@safisiol.org.ar](mailto:mini-reviews@safisiol.org.ar)

---

## **Advertising:**

For details, rates and specifications contact the Managing Editor at the Journal address e-mail:  
[mini-reviews@safisiol.org.ar](mailto:mini-reviews@safisiol.org.ar)

---

The "Sociedad Argentina de Fisiología" is a registered non-profit organization in Argentina.  
(Resol. IGJ 763-04)

---

## ***NO IN THE PARACRINE CONTROL OF THE PROXIMAL CONVOLUTED TUBULE OF THE RAT.***

**Carlos Amorena.**

**CESyMA-ECyT – Universidad Nacional de General San Martín.**

**camorena@unsam.edu.ar**

### **Proximal convoluted tubule reabsorption<sup>1</sup>.**

PCT is the main responsible for bulk reabsorption of solutes and volume in the nephron. Almost 80% of the glomerular filtrate is reabsorbed from the lumen of the PCT through the peritubular capillaries. Considering an individual of 70 Kg this amounts up to 140L of water and almost 0.5 Kg of Na<sup>+</sup> reabsorbed per day; almost three times the total body water content and ten times the total body Na content. The whole process of Na<sup>+</sup> reabsorption can be divided in three sequential steps: the first one driven mostly by the electrochemical gradient for sodium at the apical membrane, the second, which is pivotal and responsible for O<sub>2</sub> consumption of the kidney, involves the activity of the Na<sup>+</sup>-K<sup>+</sup> ATPase located at the basolateral membrane which extrudes Na<sup>+</sup> out of the cytosol, and the third one, due to solvent drag, occurring through the endothelial cells of capillaries, consequence of the Starling's equilibrium forces between the peritubular interstitium and the peritubular capillary lumen. Indeed, as a result of glomerular filtration the balance between oncotic pressure and hydrostatic pressure favors the net flow of volume from the interstitium through the capillary including the solutes present in the reabsorbed fluid.

### **Glomerulotubular balance.**

A very delicate equilibrium occurs between the amounts of ultrafiltrate formed in the Bowman's capsule and the total volume reabsorbed in the PCT. In fact, the fractional amount of volume as well as Na<sup>+</sup> reabsorbed in the PCT remains a constant fraction of the filtered values, around 70%. This relationship is known as GTB and, as indicated above, physical forces account for an important part of the maintenance of this equilibrium although factors, like Na<sup>+</sup> load and tubular fluid velocity among others have been implied in this process (1, 2, 3). The fall in the hydrostatic pressure in the efferent arteriole accompanies the increase of the oncotic pressure in the blood emerging from the glomerulus as a result of the ultrafiltration process. The water reabsorbed by the epithelium is incorporated to the circulation as a consequence of the difference between these pressures, and the solutes are included by solvent drag in the volume flux between the interstitium and the capillary lumen. Equation 1 shows the relationship between forces favoring and opposing volume flux through the endothelium of the peritubular capillaries,

$$J_v = \kappa(P h_i - P h_c + \pi_c - \pi_i) \quad \text{Eq. 1}$$

where  $\kappa$  is the hydraulic conductivity of the endothelium including the area, and  $i$  and  $c$  stands for interstitial and capillary hydraulic ( $P h$ ) and oncotic ( $\pi$ ) pressures respectively. The simultaneous determination of these driving forces allows for an analysis of the net

---

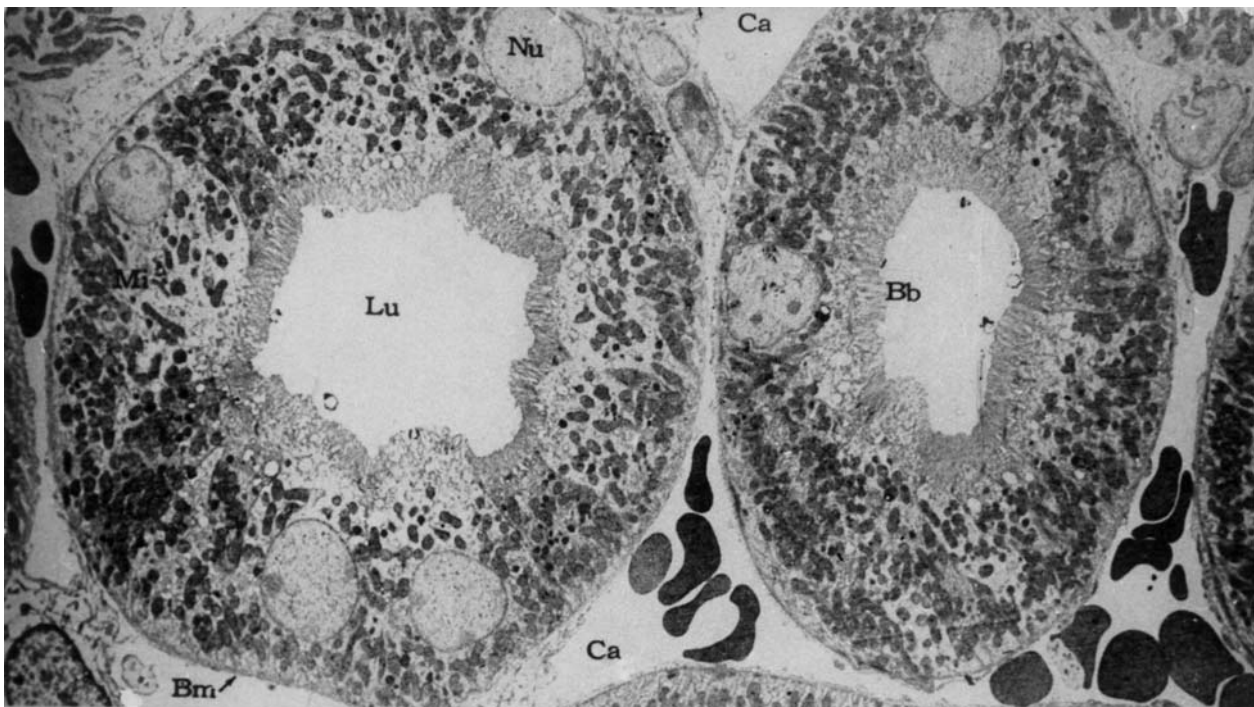
<sup>1</sup> **Glossary.** PCT: Proximal convoluted tubule, GTB: Glomerulotubular balance, NO: Nitric oxide, CCh: Carbamylcholine, BK: Bradykinin, NLA: Nitro-L-Arginine, L-NAME: Nitro-L-Arginine Methyl Ester, GB: Glibenclamide, NBCl: Na-HCO<sub>3</sub>-Cl cotransporter SNP: Sodiumnitroprusside.  $\eta$ : viscosity.

pressure favoring fluid absorption or filtration. Changes in the rate of glomerular filtration imply changes in the relative pressures operating in the postglomerular capillary bed. Indeed, these physical factors, as stated above, account for the maintenance of the glomerulotubular balance. Changes in glomerular filtration rate will also induce tubular modifications affecting the absolute proximal reabsorption (1, 2, 4).

In this review I will consider the putative role of a paracrine mechanism operating in the structure composed by the endothelium of the peritubular capillaries and the epithelium of the PCT, particularly to the effect of NO<sup>•</sup> on PCT function.

First, we must consider the close relationship between the endothelial cells and the basolateral membrane of the PCT cells.

As can be seen in the **figure 1** only a thin layer of basement membrane from both types of cell separates the endothelium from the epithelium and this anatomical arrangement facilitates the interaction between them.



**Figure 1.** Lu: Proximal tubular lumen. Nu: Nucleus. Bm: Basolateral membrana. Bb: Brush border. Ca: Capillary.

### **Hemodynamic changes due to NO<sup>•</sup>.**

The observation, by Furchgott and Zawadzki, in 1980 that a factor of short average life, released by endothelial cells accounted for vasodilatation lead not only to a Nobel Prize but also to extensive research in several areas of the biological sciences looking for the role of NO<sup>•</sup> in the control of different process, from immune response to neural differentiation. Nitric Oxide, produced in endothelial cells in response to different agonists like CCh, BK, ATP, and shear stress, acts in a paracrine way on the underlying smooth muscle cells of the vascular tree, stimulating the soluble guanylate cyclase and the formation of cGMP (4, 5, 6)

Experiments with nitro-L-arginine (NLA) and N-nitro-L-arginine methyl ester (L-NAME) have demonstrated that NO participates in the control of normal vascular tone in the kidney (7). Intra-arterial infusion of NLA in anaesthetized dogs increase renal vascular resistance 50% and reduces renal blood flow, 25% (8). Renal cortical NO activity increases steeply and linearly within the autoregulatory range, with increments in perfusion pressure. It has been suggested that it results from increased endothelial cell shear stress (9).

### **NO<sup>•</sup> and tubular transport.**

It is well known that the distal nephron is responsible for the fine tuning of final Na excretion. There are data showing that pressure natriuresis depends on decreased tubular sodium reabsorption through an amiloride sensitive transport process, implicating the distal tubule as an important site of NO<sup>•</sup> action. This is in agreement with extensive data showing the inhibitory effect of NO<sup>•</sup> on distal Na reabsorption (10).

The effect of NO<sup>•</sup> on salt and water transport in the proximal tubule, which is responsible for reabsorbing 70% of filtered sodium and water, are controversial. Whilst some studies suggest that NO<sup>•</sup> inhibits proximal tubule transport, others conclude that NO<sup>•</sup> stimulates proximal tubule transport. NOS activity measured as the conversion of L-[<sup>3</sup>H]arginine to L-[<sup>3</sup>H]citrulline and/or accumulation of NO<sup>•</sup> end-metabolites NO<sub>2</sub> or NO<sub>2</sub>/NO<sub>3</sub> was detected in isolated rat proximal tubules, primary cultures of rat or human proximal tubule cells, and proximal tubule cell lines (11, 12, 13). The mRNA for soluble guanylate cyclase, which mediates the effect of NO<sup>•</sup>, seems to be more abundant in the proximal tubule than in most other tubular segments (14). Indeed, NO<sup>•</sup> appears to modulate several types of transporters, from ionic channels through modulation of Na<sup>+</sup>-K<sup>+</sup>-ATPase, Na<sup>+</sup>/H<sup>+</sup> exchangers, and also paracellular permeability of opossum kidney cells (11, 15, 16, 17). Wang reported a biphasic effect of NO<sup>•</sup> on proximal tubular transport of sodium and bicarbonate (15). An intravenous bolus injection of L-NAME followed by addition of L-NAME to the luminal perfusate modestly decreased proximal tubular fluid reabsorption and bicarbonate reabsorption. When 1 μM sodium nitroprusside (SNP) or S-nitroso-N-acetylpenicillamine, both of which are NO<sup>•</sup> donors, was added to the luminal perfusion solution, in proximal convoluted tubules, both were increased by 30~50%. However, proximal tubular volume and bicarbonate decreased by 50~70% with 1mM SNP. On the other hand, perfusion of the tubule lumen with dibutyl cGMP increased net proton flux. Agents that elicit NO<sup>•</sup> production, like BK, Cch, and ATP increase proton flux when added to the peritubular capillary perfusate. These effects were blocked by L-NAME. Bradykinin increased cGMP content of isolated proximal convoluted tubules, but only if they were co-incubated with endothelial cells (17, 18).

Basal NO<sup>•</sup> production stimulates Na<sup>+</sup>/H<sup>+</sup> exchange in proximal tubules of rats (16, 17, 18), whereas higher NO<sup>•</sup> concentrations from exogenous sources inhibit Na<sup>+</sup>/H<sup>+</sup> exchange and Na<sup>+</sup> reabsorption. It is important to note that doses of the SNP inhibit Na<sup>+</sup>-K<sup>+</sup> ATPase and Na<sup>+</sup> reabsorption in proximal cells in the range of 0.4-1 mM which are substantially higher than the dose shown to stimulate proximal tubular reabsorption in vivo (19, 20). Interestingly, cholinergic agents increase the activity of the renal Na<sup>+</sup>-HCO<sub>3</sub><sup>-</sup> cotransporter, this effect is mimicked by SNP and blocked by L-NAME (21). The increase in the Na<sup>+</sup>/H<sup>+</sup> activity due to NO is consistent with this effect.

Another important agonist of NO<sup>•</sup> formation in the vascular system is shear stress (22, 23). A viscous fluid (blood) moving along a solid boundary will undergo a shear stress on that boundary. In this condition the fluid has zero velocity relative to the boundary, but at some distance from the boundary the flow speed must equal that of the fluid. The shear stress is imparted onto the boundary (in our case the apical membrane of the endothelial cells) as a result of this loss of velocity and can be expressed as (Eq 2)

$$ss = \eta \frac{dv}{dy} \quad \text{Eq. 2}$$

where  $\eta$  is the viscosity of the fluid,  $v$  is the velocity of the fluid along the boundary, and  $y$  is the distance from the boundary. Thus we can consider shear stress as an agonist of NO production like like BK, Cch etc. This point is important since as result of glomerular filtration the viscosity of the emerging blood in the efferent arteriole must increase. The increased  $\eta$  results from the increase of hematocrit (Htc)—with a filtration fraction of 0.3 Htc should raise from 0.45 to 0.64—as well as of the protein concentration. From a physiological point of view, increased  $\eta$  at the efferent arteriole would justify the generation of NO<sup>•</sup> that is able to affect functionally epithelial structures in the surroundings of the arteriole.

Changes in the viscosity of peritubular perfusate modify the reabsorption of bicarbonate in proximal convoluted tubules of rat's kidney. The  $\eta$ -induced increase in PCT H<sup>+</sup> flux was blocked by L-NNA, suggesting that it depends on NO release. L-NNA not only abolished the effect of dextran, but also inhibited H<sup>+</sup> flux below the control value. These results suggest that there is a basal NO-dependent H<sup>+</sup> flux. Moreover, the inhibitory effect of L-NNA on H<sup>+</sup> flux stimulated by high  $\eta$  was abolished by L-arginine and by P<sub>2</sub>-purinoreceptor blockade with Suramin and RB2 (20). It has been shown that ATP is released from the endothelium by shear stress (22, 23). Therefore, changes in  $\eta$  could modify NO<sup>•</sup> synthesis through ATP release. These results strongly suggest that NO produced by increased  $\eta$  stimulates the synthesis of cGMP in tubular epithelial cells, activating the Na<sup>+</sup>/H<sup>+</sup> exchanger, and thus coupling filtered Na<sup>+</sup> load to proximal tubule Na<sup>+</sup> and water reabsorption. The present results suggest that a mechanism of this type could contribute, in addition to the Starling forces in the peritubular capillaries, to the control of GTB.

Given the structure of the peritubular capillaries net, it would not be strictly proper to consider blood flow to be Newtonian, thus the concept of viscosity turns to be rather misapplied. The apparent viscosity of blood in glass tubes declines with decreasing diameter (Fåhræus-Lindqvist effect) and exhibits a distinctive minimum at 6–7  $\mu\text{m}$ . However, flow resistance in vivo in small vessels is substantially higher than predicted by in vitro viscosity data. However, Lipowsky et al. (24) showed that the apparent blood viscosity is much higher in relatively straight, unbranched microvessels than in glass tubes with corresponding inner diameters.

### **Effect of NO<sup>•</sup> on basolateral membrane voltage.**

Finally, an additional effect of NO<sup>•</sup> on proximal convoluted tubules cells has been documented. It has been reported that NO<sup>•</sup> affects the activity of K<sup>+</sup> channels in the distal nephron segments, such as cortical collecting duct and thick ascending limb (25, 26).

Nakamura et al showed that NO<sup>•</sup> modulates the activity of an inwardly rectifying K<sup>+</sup> channel in cultured human proximal tubule cells (27). These authors also show that the stimulatory effects of NO<sup>•</sup> donors is abolished by a PKG-specific inhibitor and concluded that NO<sup>•</sup> stimulated channel activity through the activation of soluble guanylate cyclase, consequent elevation of intracellular cGMP, and PKG-mediated phosphorylation. With micropuncture experiments we obtained evidence that agonists of NO<sup>•</sup> formation at the peritubular capillaries affects basolateral membrane potential ( $V_{blm}$ ). Changes in the  $\eta$  of the peritubular perfusate modified membrane potential at the basolateral membrane of PCT epithelial cells. As PCT epithelium is leaky, changes in  $V_{blm}$  affect also the apical membrane potential and thus the electrical gradient potential for Na<sup>+</sup> uptake. Also, BK, a chemical agonist, hyperpolarized the cells and both increased viscosity and BK effects were diminished by I-NAME. The effects of high  $\eta$  seem to involve the ATP sensitive K<sup>+</sup> conductance as GB abolishes the effect of increased viscosity (28). As it was discussed above, there is evidence that NO<sup>•</sup> stimulates Na<sup>+</sup>-H<sup>+</sup> exchanger at the apical membrane of PCT. This would lead to an incremented Na<sup>+</sup>-K<sup>+</sup> ATPase activity. There is evidence that the activity of the Na<sup>+</sup>-K<sup>+</sup> ATPase is linked to the K<sup>+</sup> conductance (29). An increase in Na<sup>+</sup>-K<sup>+</sup> ATPase activity from an increase in Na<sup>+</sup> transport at the apical membrane domain, could lead to a decrease in ATP level and, consequently, to an increase in K<sup>+</sup> conductance. The NBC1 activity induces a depolarizing current at the basolateral membrane of the PCT (30). The effects of shear stress enhancing Na<sup>+</sup>/H<sup>+</sup> exchange at the apical membrane and HCO<sub>3</sub><sup>-</sup> reabsorption through the NBC1 would generate a depolarizing current at the basolateral membrane. The hyperpolarizing effect at the basolateral membrane could counterbalance the depolarizing current, preserving the driving force for Na<sup>+</sup> from the lumen. The effects of HCO<sub>3</sub><sup>-</sup> transport at the basolateral membrane lead to a decrease in the electrical gradient for Na<sup>+</sup> across the apical membrane. The increase in gKATP must counterbalance the depolarizing effect of an increase in HCO<sub>3</sub><sup>-</sup> transport across the basolateral membrane.

All these results suggest the presence of a delicate equilibrium among these conductive transport processes in PCT that could be, at least in part, modulated naturally by glomerular filtration rate. The analysis of the mechanism as a whole seems to indicate that different control systems concur to damp fluctuation due to changes in GFR, i.e., tubuloglomerular feedback, thus maintaining the proximal Na<sup>+</sup> and water reabsorption constant in spite of changes in Na<sup>+</sup>-filtered load. From a functional point of view, it looks like changes in shear stress through NO<sup>•</sup> release modulate PCT epithelial cells function, synchronizing different transport mechanisms affecting the reabsorption process and coupling GFR with PCT reabsorption.

## **REFERENCES**

1. Mathisen O, Monclair T, Kiil F. Factors limiting proximal tubular reabsorption at high glomerular filtration rate. *Scand J Clin Invest* 38:573, 1978.
2. Haberle DA, Shiigai TT, Maier G, et al. Dependency of proximal tubular fluid transport on the load of glomerular filtrate. *Kidney Int* 20:18, 1981.
3. Imai M, Seldin DW, Kokko JP. Effect of perfusion rate on the fluxes of water, sodium, chloride and urea across the proximal convoluted tubule. *Kidney Int* 11:18, 1977.
4. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle. *Nature* 288:373-376, 1980.
5. Ignarro LJ, Buga GM, Wood KS, et al. Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. *Proc Natl Acad Sci USA* 84:9265, 1987.

6. Palmer RMJ, Ferridge AG, Moncada S. Nitric oxide release accounts for the biologic activity of endothelium-derived relaxing factor. *Nature* 327:524, 1987.
7. Majid DS, Navar LG. Nitric oxide in the control of renal hemodynamics and excretory function. *Am J Hypertens.* 14(6 Pt 2):74S-82S, 2001 Jun.
8. Majid DS, Navar LG. Suppression of blood flow autoregulation plateau during nitric oxide blockade in canine kidney. *Am J Physiol.* 1992 Jan; 262(1 Pt 2):F40-6, 1992.
9. Majid DS, Godfrey M, Grisham MB, Navar LG. Relation between pressure natriuresis and urinary excretion of nitrate/nitrite in anesthetized dogs. *Hypertension.* 25(4 Pt 2):860-5, 1995 Apr.
10. Ortiz PA, Garvin JL. Role of nitric oxide in the regulation of nephron transport. *Am J Physiol Renal Physiol* 282: F777-F784, 2002.
11. Guzman NJ, Fang MZ, Tang SS, Ingelfinger JR, Garg LC. Autocrine inhibition of Na<sup>+</sup>/K<sup>(+)</sup>-ATPase by nitric oxide in mouse proximal tubule epithelial cells. *J Clin Invest* 95: 2083-2088, 1995.
12. McLay JS, Chatterjee P, Nicolson AG, Jardine AG, McKay NG, Ralston SH, Grabowski P, Haites NE, MacLeod AM, Hawksworth GM. Nitric oxide production by human proximal tubular cells: a novel immunomodulatory mechanism? *Kidney Int* 46: 1043-1049, 1994.
13. Yu L, Gengaro PE, Niederberger M, Burke TJ, Schrier RW. Nitric oxide: a mediator in rat tubular hypoxia/reoxygenation injury. *Proc Natl Acad Sci USA* 91: 1691-1695, 1994.
14. Terada Y, Tomita K, Nonoguchi H, Marumo F. Polymerase chain reaction localization of constitutive nitric oxide synthase and soluble guanylate cyclase messenger RNAs in microdissected rat nephron segments. *J Clin Invest* 90: 659-665, 1992.
15. Liang M, Knox FG. Nitric oxide enhances paracellular permeability of opossum kidney cells. *Kidney Int* 55: 2215-2223, 1999.
16. Wang, T. Nitric oxide regulates HCO<sub>3</sub><sup>-</sup> and Na<sup>+</sup> transport by a cGMP-mediated mechanism in the kidney proximal tubule. *Am J Physiol Renal Physiol* 272: F242-F248, 1997.
17. Diaz-Sylvester P, Mac Laughlin M, Amorena C. Peritubular fluid viscosity modulates H<sup>+</sup> flux in proximal tubules through NO release. *Am J Physiol Renal Physiol* 280: F239-F243, 2001.
18. Amorena C, Castro AF. Control of proximal tubule acidification by the endothelium of the peritubular capillaries. *Am J Physiol Regul Integr Comp Physiol* 272: R691-R694, 1997.
19. Liang M, Knox FG. Nitric oxide reduces the molecular activity of Na<sup>+</sup>-K<sup>+</sup>-ATPase in opossum kidney cells. *Kidney Int* 56: 627-634, 1999.
20. Roczniak A, Burns KD. Nitric oxide stimulates guanylate cyclase and regulates sodium transport in rabbit proximal tubule. *Am J Physiol Renal Fluid Electrolyte Physiol* 270: F106-F115, 1996.
21. Ruiz OS, Qiu YY, Cardoso LR, Arruda JA. Role of nitric oxide and intracellular calcium. *Miner Electrolyte Metab.* 25(3):171-7, 1999 May-Jun.
22. Burnstock, G. Release of vasoactive substances from endothelial cells by shear stress and purinergic mechanosensory transduction. *J Anat* 194: 335-342, 1999.
23. Milner P, Kirkpatrick KA, Ralevic V, Toothill V, Pearson J, Burnstock G. Endothelial cells cultured from human umbilical vein release ATP, substance P and acetylcholine in response to increased flow. *Proc Soc Lond B Biol Sci* 241: 245-248, 1990.
24. Lipowsky HH, Usami S, Chien S. In vivo measurements of "apparent viscosity" and microvessel hematocrit in the mesentery of the cat. *Microvasc Res* 19: 297-319, 1980.
25. Lu M, Giebisch G, Wang W. Nitric oxide links the apical Na<sup>+</sup> transport to the basolateral K<sup>+</sup> conductance in the rat cortical collecting duct. *J Gen Physiol* 110: 717-726, 1997.
26. Lu M, Wang X, Wang W. Nitric oxide increases the activity of the apical 70-pS K<sup>+</sup> channel in TAL of rat kidney. *Am J Physiol Renal Physiol* 274: F946-F950, 1998.
27. Nakamura K, Hirano J, Kubokawa M. Regulation of an inwardly rectifying K<sup>+</sup> channel by nitric oxide in cultured human proximal tubule cells. *Am J Physiol Renal Physiol.* 287(3):F411-7, 2004 Sep.
28. Lopardo M, Diaz-Sylvester P, Amorena C. The effect of shear stress on the basolateral membrane potential of proximal convoluted tubule of the rat kidney. *Pflugers Arch* 454:289-295, 2007.
29. Muto S, Asano Y, Wang W, Seldin D, Giebisch G. Activity of the basolateral K<sup>+</sup> channels is coupled to the Na<sup>+</sup>-K<sup>+</sup>-ATPase in the
30. Seki G, Coppola S, Yoshitomi K, Burckhardt BC, Samarzija I, Muller-Berger S, Fromter E. On the mechanism of bicarbonate exit from renal proximal tubular cells. *Kidney Int* 49(6):1671-1677, 1996.